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Abstract Journal

CD/5-FC 系统对结肠癌细胞的杀伤作用

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Killing effect of CD/5-FC system on human colon cancer cell lines SW 480 and LoVo

Cheng-Jin Li, Qing-Jiu Ma, Da-Nian Lai, Jiang-Guo Lu, Xiao-Jun Wang, Qing Wang, Bo-Rong Pan, Yong-Zhong Wu, Jin-Mao Li

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Abstract

AIM:To investigate the killing effect of carcinoembryonic antigen (CEA) and tissue-specific cytosine deaminase (CD)/5-fluorocytosine (5-FC) system on human colorectal carcinoma cell lines LoVo and SW480 *in vitro*.

METHODS:Recombinant retroviral vector G1CEACDNa was constructed, in which the CD gene was controlled under the CEA promoter, and retroviral vector pCD2 were introduced through liposome technique respectively to the human colorectal carcinoma cell lines LoVo and SW480. Expression of CEA was high and low in both the cell lines respectively. The cells were selectively cultured in G418. The proliferative colonies were treated with 5-FC.

RESULTS:After the transfection, LoVo-CEACD cells and LoVo-CD cells were more sensitive to 5-FC than their parental cells ($P < 0.01$, $t = 5.688$, $n = 9$; $P < 0.01$, $t = 3.136$, $n = 9$), and SW480-CEACD cells and SW480-CD cells were more sensitive than their parental cells as well ($P < 0.01$, $t = 3.437$, $n = 9$; $P < 0.01$, $t = 3.516$, $n = 9$). Furthermore, the LoVo-CEACD cells were more sensitive to 5-FC than the LoVo-CD cells ($P < 0.05$, $t = 2.183$, $n = 9$) while the SW480-CEACD cells were less sensitive than SW480-CD cells. The LoVo-CEACD cells displayed a higher anti-tumor effect than SW480-CEACD cells *in vitro*. The bystander effect in all cells transfected with CD gene were observed in this study.

CONCLUSION:The CEA tissue-specific CD/5-FC system

displays an obvious targeting anti-tumor effect on human colorectal carcinoma cell lines LoVo and SW480, but the killing effect on the LoVo-CEACD cells is higher than that on the SW480-CEACD cells *in vitro*.

Li CJ, Ma QJ, Lai DN, Lu JG, Wang XJ, Wang Q, Pan BR, Wu YZ, Li JM. Killing effect of CD/5-FC system on human colon cancer cell lines SW 480 and LoVo. Shijie Huaren Xiaohua Zazhi 2003;11(5):535-539

摘要

目的:探讨组织特异性胞嘧啶脱氨酶(cytosine deaminase, CD)/5-氟胞嘧啶(5-fluorocytosine, 5-FC)系统对不同分泌癌胚抗原(carcinoembryonic antigen, CEA)的大肠癌细胞 LoVo 和 SW480 的靶向杀伤作用。

方法:脂质体法将 CEA 基因顺式转录调控序列(TRS)驱动 CD 基因的组织特异性逆转录病毒载体 G1CEACDNa 及非组织特异性逆转录病毒载体 pCD2 分别转入大肠癌细胞 LoVo 和 SW480, 以 G418 筛选阳性克隆扩增后给予前药 5-FC 进行敏感试验。

结果:LoVo-CEACD 及 LoVo-CD 比 LoVo 对 5-FC 的敏感性明显提高($P < 0.01$, $t = 5.688$, $n = 9$; $P < 0.01$, $t = 3.136$, $n = 9$), SW480-CEACD 及 SW480-CD 比 SW480 对 5-FC 的敏感性明显提高($P < 0.01$, $t = 3.437$, $n = 9$; $P < 0.01$, $t = 3.516$, $n = 9$), LoVo-CEACD 比 LoVo-CD 对 5-FC 的敏感性明显增强($P < 0.05$, $t = 2.183$, $n = 9$), 而 SW480-CEACD 对 5-FC 的敏感性小于 SW480-CD, SW480-CEACD 对前药 5-FC 的敏感性低于 LoVo-CEACD ($P < 0.05$, $t = 2.504$, $n = 9$), 转 CD 基因之 LoVo 和 SW480 细胞体外实验均可观察到明显的旁观者效应。

结论:组织特异性 CD/5-FC 系统对 LoVo 和 SW480 细胞均有明显的靶向杀伤效果, 但对 SW480 细胞的杀伤作用小于 LoVo 细胞。

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<http://www.wjgnet.com/1009-3079/11/535.htm>

0 引言

CD/5-FC 系统作为前药转换基因系统之一, 近年来对其在肿瘤治疗中的作用进行了广泛研究^[1-4], 已进行了少量 期临床实验^[1,5-8]. 研究表明^[9], 癌胚抗原(CEA)转录调控序列可控制 CD 基因在 CEA 阳性的大肠癌组织中高效表达, 在前药 5-FC 作用下, 产生选择性杀伤

肿瘤细胞的作用.然而,CEA启动子控制的CD基因对低表达CEA的大肠癌细胞是否具有靶向性杀伤作用?为此,我们进行了初步研究.

1 材料和方法

1.1 材料 含CD基因逆转录病毒载体pCD2由美国Anderson癌症研究中心的Mullen博士惠赠;CEA启动子调控CD基因表达的逆转录病毒载体G1CEACDNa由第二军医大学长海医院崔龙教授惠赠;大肠癌LoVo细胞株由上海医科大学瑞金消化外科研究所提供;大肠癌SW480细胞株由第四军医大学动物研究所提供;感受态菌由第四军医大学遗传与发育教研室制备;脂质体Iopfectamine™ 2 000 (LF 2 000)为Invitrogen公司产品;G 418, DMEM为Gibco公司产品;5-FC, MTT为Sigma公司产品;RPMI 1640培养基,胎牛血清, BamH 酶, EcoR 酶, Sal 酶为Promega公司产品;大量柱离心式质粒抽提纯化试剂盒及小量柱离心式组织和细胞基因组DNA抽提试剂盒为上海华舜公司产品;PCR扩增Premix Taq™ 酶为Biotech公司产品,CD基因引物由Biotech公司合成,引物序列(扩增产物1.5 kb)为:正义链5'-ATAGAATTCAGGCTAACAA TGTCGAATTAACGCTT-3',反义链5'-TATGGATCCTCAACGTTTGTAAATCCATGGCTT-3'.

1.2 方法

1.2.1 质粒扩增和酶切鉴定 将质粒pCD2, G1CEACDNa在感受态中大量扩增后,按质粒抽提试剂盒说明书提取并纯化质粒,主要步骤:在细菌沉淀中加入P1液3.5 mL,振荡后加入P2液3.5 mL并混匀,室温静置4 min,加入PN液5 mL并混匀,12 000 g离心30 min,将上清液移入黏附柱,离心2 min,加入W1液12 mL,静置1 min,离心2 min,在黏附膜中央加入T1液500 μL,室温静置2 min后,离心5 min.质粒pCD2用BamH及EcoR酶切鉴定,质粒G1CEACDNa用Sal酶切鉴定,用20 g/L琼脂糖凝胶电泳.

1.2.2 大肠癌细胞的转染 在24孔培养板中分别接种 3×10^5 个SW480、LoVo细胞,待细胞生长至95%融合时,用无血清RPMI 1640培养基轻洗细胞并用之置换加宁L(含1 μg质粒DNA和2.5 μL LF 2 000)培养24 h,以含400 mg/L G 418及100 mL/L胎牛血清的RPMI 1 640培养基传代培养转染细胞,选择培养14 d,其中原传代2次,筛选抗性克隆并扩增.

1.2.3 细胞基因组DNA的提取及PCR检测 按细胞DNA提取试剂盒说明书操作,主要步骤如下:将 10^7 细胞移入离心管中,10 000 g离心10 s,弃上清,加入RNase A 20 μL和DT液200 μL,混匀,65℃温浴5 min,加入DL液400 μL和Proteinase K 25 μL并混匀后65℃温浴20 min,离心5 min,将上清液移入另一离心管中,加入异丙醇200 μL混匀后,取样品液至黏附柱中,离心30 s,加入W1液500 μL,静置1 min,离

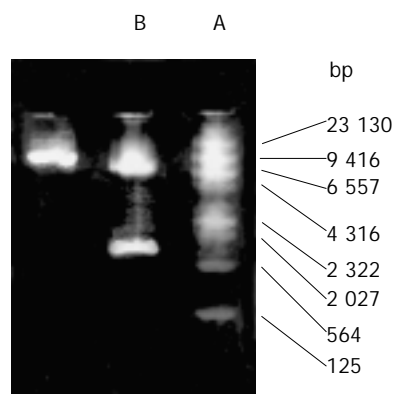
心30 s,加入W1液500 μL,离心30 s,在黏附柱中央加入T1液100 μL,65℃静置5 min,离心1 min, -20℃保存. PCR用50 μL反应体系,反应条件:94℃, 3 min, 94℃, 30 s, 60℃, 30 s, 72℃, 40 s, 72℃, 10 min, 40个循环,PCR产物用20 g/L琼脂糖凝胶电泳.

1.2.4 前药5-FC对转基因大肠癌细胞的杀伤作用 将转染G1CEACDNa、pCD2之LoVo、SW480细胞(分别命名为:LoVo-CEACD, LoVo-CD, SW480-CEACD, SW480-CD)及未转基因之LoVo、SW480细胞以 5×10^4 个/孔接种到96孔细胞培养板中,接种同时加入含各种梯度浓度的前药5-FC,每种浓度设3个复孔,同时设置对照孔及调零孔.在37℃、50 mL/L CO₂孵箱中培养,每2 d换液1次,第8天去除培养液,以MTT法测定活细胞比率并计算杀伤率:加入MTT 20 μL(5g/L), 37℃孵育4 h后弃上清加入二甲基亚砷(Sigma)150 μL, 10 min后振荡,以490为测定波长,上酶标仪测定吸光度值(A).根据下列公式计算细胞存活率:存活率=实验组A值/对照组A值 $\times 100\%$.将转CD基因SW480, LoVo细胞与未转基因之SW480、LoVo细胞分别以100, 80, 50, 30, 20, 10, 0之比率(%)混合接种至24孔板中,每种混合细胞设3个复孔,同时设置对照孔及调零孔,以含2.0 mmol/L 5-FC的RPMI 1 640完全培养基中培养8 d后,以MTT法检测活细胞比率并计算旁观者效应杀伤率.

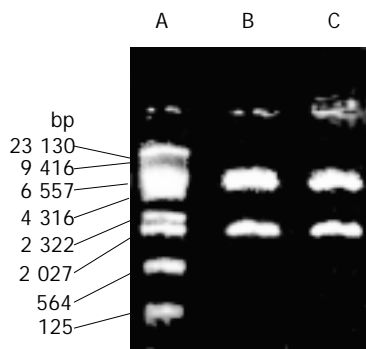
统计学处理 在SPSS软件中输入药敏实验中各浓度前药下每组细胞生存率,作线图并计算IC₅₀,采用未配对计量资料的t检验进行统计学处理.

2 结果

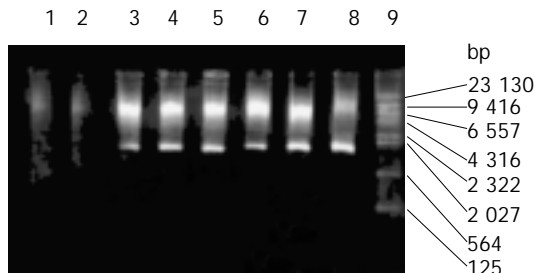
2.1 目的基因整合到靶细胞中 质粒pCD2用BamH及EcoR酶切可见1.5 kb酶切片段(图1),质粒G1CEACDNa用Sal酶切可见1.9 kb酶切片段(图2).转pCD2及G1CEACDNa之SW480、LoVo细胞PCR产物电泳均可见1.5 kb片段(图3).



A. λ DNA Hind Marker; B. pCD2 plasmid.
图1 pCD2质粒酶切鉴定.



A. λ DNA Hind Marker; B,C: G1CEACDNa plasmid.
图2 G1CEACDNa 质粒酶切鉴定.



1: SW480; 2: LoVo; 3: SW480-CEACD; 4, 5: SW480-CD; 6: LoVo-CD
7, 8: LoVo-CEACD; 9: λ DNA Hind Marker.
图3 PCR 验证 CD 基因转染 LoVo 和 SW480 细胞.

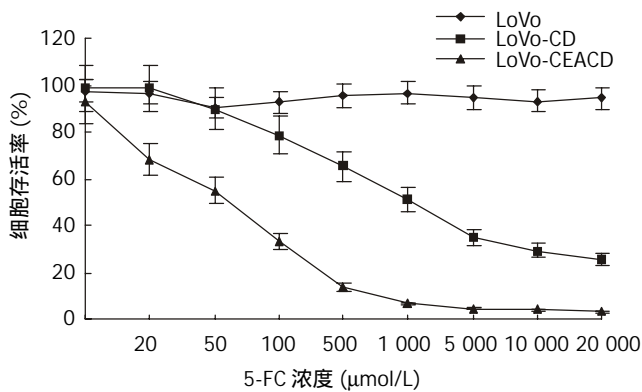


图4 前药 5-FC 对转 CD 基因的 LoVo 细胞的杀伤作用.

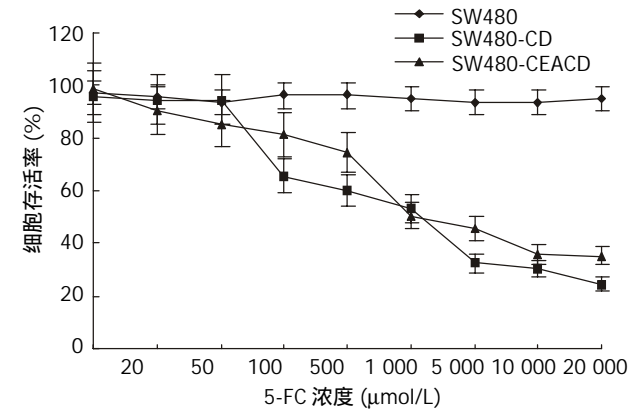


图5 前药 5-FC 对转 CD 基因的 SW480 细胞的杀伤作用.

2.2 前药 5-FC 对转基因大肠癌细胞 LoVo、SW480 的杀伤作用 LF2 000 转染 pCD2、G1CEACDNa, 以含 400 mg/L G418 的 1 640 培养基选择培养 14 d, 对照组细胞在 6 d 后

全部死亡. 5-FC 对转基因大肠癌细胞 LoVo, SW480 有杀伤作用(图 4, 5), 转 CD 基因之 LoVo, SW480 细胞对 5-FC 的敏感性明显提高, 转 G1CEACDNa 之 LoVo 细胞较转 pCD2 之 LoVo 细胞对 5-FC 的敏感性明显增强, 其 IC_{50} 分别为 0.1 mmol / L 和 0.8 mmol / L, 而转 G1CEACDNa 之 SW480 细胞对 5-FC 的敏感性小于转 pCD2 之 SW480 细胞, 其 IC_{50} 分别为 1.0 mmol / L 和 0.8 mmol / L. 细胞的杀伤率与转基因细胞的比例并不一致, 当转基因细胞达到 30 % 时, 对混合细胞的杀伤作用就与 100% 转基因细胞的杀伤作用相似(旁观者效应, 表1).

表1 旁观者效应细胞杀伤率 (%) , $\bar{x} \pm s$ $n=3$)

细胞	转基因细胞比率						
	0	10	20	30	50	80	100
LoVo-CD	0	31.5	44.5	58.2	61.3	63.6	71.5
LoVo-CEACD	0	45.6	79.4	88.5	91.3	92.8	96.5
SW480-CD	0	32.5	41.3	53.5	60.2	61.2	63.2
SW480-CEACD	0	33.3	41.3	52.0	53.6	55.5	59.3

3 讨论

胞嘧啶脱氨基酶(cytosine deaminase, CD)基因是来源于某些细菌和真菌的一种自杀基因, 其编码的 CD 酶能将对真核细胞相对无毒的 5- 氟胞嘧啶(5-FC)转换成细胞毒性化疗药 5- 氟尿嘧啶(5-FU), 抑制细胞 RNA 和 DNA 的合成而致细胞死亡^[10]. 5-FU 作为结肠癌的一线化疗药物, 对机体正常组织的毒副作用限制了其在结肠癌化疗的临床应用^[11-13], 靶向基因治疗可将治疗基因在肿瘤组织中特异地表达, 使正常组织免受损害. CEA 基因属于组织特异性表达基因, 利用 CEA 基因的转录调控序列来调控 CD 基因的表达, 可特异性杀死 CEA 阳性分泌的大肠癌细胞^[14-16], 对结肠癌肝转移也有特异性治疗作用及旁杀伤效应^[17-21], 对结肠癌的腹膜转移亦能进行有效治疗^[22]. 临床资料显示, 虽然 CEA 作为结肠癌疗效及转移的监测指标有其特异性^[23], 但在大肠癌患者中, 近 2/3 血清 CEA 正常^[24], Dukes A 期患者血清 CEA 水平仅 $5.3 \pm 1.8 \mu\text{g/L}$ ^[25], 表明临床有相当一部分大肠癌患者其 CEA 表达水平较低. 我们所采用的 LoVo 和 SW480 分别是高表达 CEA 和低表达 CEA 的人大肠癌细胞株, CEA 分泌水平分别为 49 fg/cell 和 3.1 fg/cell.

本研究用逆转录病毒载体 G1CEACDNa 及 pCD2 分别转染 LoVo 细胞和 SW480 细胞, G1CEACDNa 由组织特异性 CEA 基因顺式转录调控序列(TRS)调控 CD 基因的表达, pCD2 为非组织特异性 CD 基因逆转录病毒载体, 我们用脂质体 LF2000 直接转染大肠癌细胞, 经 G418 选择培养 14 d 后, 用前药 5-FC 进行实验. 结果显示, 目的基因均成功整合到靶细胞中, LoVo-CEACD 及 LoVo-CD 细胞比未转基因之 LoVo 细胞对 5-FC 的敏

感性明显提高($P < 0.01$, $t = 5.688$, $n = 9$; $P < 0.01$, $t = 3.136$, $n = 9$), SW480-CEACD及SW480-CD细胞比未转基因之SW480细胞对5-FC的敏感性明显提高($P < 0.01$, $t = 3.437$, $n = 9$; $P < 0.01$, $t = 3.516$, $n = 9$), LoVo-CEACD较LoVo-CD细胞对5-FC的敏感性明显增强($P < 0.05$, $t = 2.183$, $n = 9$), 其 IC_{50} 提高了8倍, 而SW480-CEACD细胞对5-FC的敏感性小于SW480-CD细胞, 后者的 IC_{50} 是前者的1.25倍; 转染CEA组织特异性CD基因后, 低表达CEA的大肠癌细胞SW480对前药5-FC的敏感性低于高表达CEA的人大肠癌细胞LoVo($P < 0.05$, $t = 2.504$, $n = 9$), 后者 IC_{50} 是前者的10倍; 转G1CEACDNa及pCD2之LoVo和SW480细胞均可观察到明显的旁观者效应. 我们的实验结果表明, 用组织特异性CEA启动子调控CD基因的表达, 对高表达CEA的大肠癌细胞LoVo和低表达CEA的大肠癌细胞SW480均有明显的靶向杀伤作用, 提高了细胞对前药5-FC的敏感性, 但对SW480的杀伤作用不如对LoVo细胞的杀伤作用理想, 甚至低于普通型CD基因, 分析可能是由于肿瘤细胞CEA的低表达导致CEA启动子调控的CD基因的低表达所致, 其机制有待进一步实验证实.

靶向性和有效性是基因治疗的两大瓶颈, 前者可使正常组织细胞免受损害, 后者保证了治疗的效率. 我们的实验提示, CEA组织特异性CD/5FC系统对低表达CEA大肠癌的治疗效率将受到影响, 有研究表明, 用CEA启动子和增强子共同调节CD基因的表达, 能明显提高肿瘤细胞CD基因的表达^[26], 放疗、免疫因子与CD/5FC系统的联合应用、双自杀基因的共同转导^[27-38]等都能提高CD/5FC系统的治疗效果, 这样可以在保证治疗靶向性的前提下提高治疗效率.

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