

• 幽门螺杆菌 *H pylori* •

小鼠 *H pylori* 长期感染模型研究

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Establishment of mouse models with long-term infection of *H pylori*

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Abstract

AIM: To develop a mouse model with long-term infection of *H pylori*.

METHODS: Each mouse was inoculated with *H pylori* Sydney strain 1 (SS1). Noninfected control mice and infected mice were killed at 4, 12 and 24 wk after *H pylori* infection. A piece of gastric mucosa obtained from the posterior wall of the antrum of each mouse was used for culture of *H pylori*, rapid urease testing and histopathological detection. Serum was obtained to measure the IgG antibody level to *H pylori*.

RESULTS: After 4 wk of infection, the *H pylori* was cultured from 6 of 7 infected Balb/c mice and 5 of 7 the *H pylori*-infected C57BL/6 mice. After 12 and 24 wk of infection, all of 7 infected Balb/c and 6 of 7 the *H pylori*-infected C57BL/6 mice showed the positive results. At the different wk, the rapid urease test results were consistent with the *H pylori* culture test. At 12 wk after inoculation, chronic inflammation was observed in the pyloric mucosa by histopathological examination. At 24 wk after inoculation, lymphoid follicles were especially conspicuous in the submucosa, and they were also found in the deep portion of the mucosa.

CONCLUSION: *H pylori* SS1 can colonize easily in the glandular stomach mucosa of mouse, and the histopathological changes are similar to those of humans with *H pylori* infection.

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摘要

目的: 用 *H pylori* SS1 (Sydney strain1) 感染 C57BL/6 小鼠及 Balb/c 小鼠, 建立稳定的 *H pylori* 感染的小鼠实验动物模型。

方法: 通过灌胃的方式用 *H pylori* SS1 菌株感染二级 C57BL/6 小鼠和 Balb/c 小鼠, 灌胃后 4、12 及 24 wk 分三次处死动物, 用胃黏膜匀浆液进行细菌培养试验、尿素酶试验、胃病理组织学检查以及血清学抗体检测试验。

结果: 灌胃 4 wk 后, 处死的 7 只 Balb/c 小鼠经胃组织细菌培养发现有 6 只显示阳性结果; 处死的 7 只 C57BL/6 小鼠中有 5 只显示阳性结果。12 和 24 wk 后的实验组 7 只 Balb/c 小鼠胃组织细菌培养均为阳性, 而 7 只 C57BL/6 小鼠中有 6 只显示阳性结果。在不同阶段尿素酶检测的阳性结果与细菌培养法结果一致。实验组动物胃窦部及胃体部均出现了轻度至中度慢性活动性胃炎变化; 血清学 IgG 检查表明, 各实验组的实验动物产生了抗 *H pylori* 血清抗体; 而对照组动物的胃黏膜匀浆液经各项检测均为阴性。

结论: 成功地建立了 *H pylori* 感染的小鼠实验动物模型。

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0 引言

H pylori 感染可导致人体发生慢性活动性胃炎和胃、十二指肠的消化性溃疡等, 同时有诱发机体发生胃癌的危险^[1-5]。*H pylori* 的感染率极高, 在发达国家 50% 以上的成年人感染该菌, 发展中国家的感染率达到 60-80%, 甚至更高^[6-10]。调查表明, 90% 的胃炎和 70% 的胃溃疡由 *H pylori* 引起, 而且 *H pylori* 感染所致疾病的治疗尚未得到很好的解决^[11-14]。目前, *H pylori* 的致病机制仍然不十分清楚, 从根本上解决防治 *H pylori* 相关性疾病的首要方法是 *H pylori* 疫苗的研制和临床应用, 而建立稳定的动物模型是探讨 *H pylori* 致病机制及开展特异性免疫预防与治疗的前提和保证^[15-16]。现在建立的动物模型虽然不少, 但是方便而价廉的 *H pylori* 小鼠模型的建立还少有报道。我们通过灌胃的方式用 *H pylori* SS1 菌株感染二级 C57BL/6 小鼠和 Balb/c 小鼠, 表明 *H pylori* SS1 菌株可长期定植于 Balb/c 和 C57BL/6 小鼠胃黏膜, 并发生了胃黏膜炎症变化, 成功地建立了 *H pylori* 感染的小鼠实验动物模型, 为 *H pylori* 致病机制的研究、疫苗开发研制等奠定了基础。结果如下。

1 材料和方法

1.1 材料 二级C57BL/6小鼠及Balb/c小鼠各30只, 17-20 g, ♀; 由本院实验动物中心提供, 本所动物房饲养。H pylori SS1 (Sydney strain1)由广州中山医科大学陈曼湖教授惠赠。

1.2 方法 每种动物随机分成实验组(21只)和对照组(9只)。动物灌胃前进行8 h禁食和4 h禁水, 然后用30 mg/L NaHCO₃ 0.25 mL灌胃, 30 min后用1×10⁹ cfu/L H pylori SS1 0.5 mL灌胃, 灌后30 min恢复进食和饮水^[17]。对照动物灌服PBS。隔日连续灌服3次。末次灌胃后的4、12及24 wk分3次, 每次7只, 眼球放血并处死动物。于无菌超净台中取出处死动物的胃, 用灭菌生理盐水清洗胃中内容物, 沿胃大弯将胃黏膜纵切为两半, 其中一半置于甲醛液体中常规固定, 用于病理组织学检查; 另一半用剪刀剪碎后(剪碎前称质量), 各加生理盐水0.5 mL于研磨器中研磨至匀浆进行细菌培养计数, 并用H pylori 快速检测试剂盒进行H pylori 尿素酶检测。采用空肠弯曲菌琼脂基础培养基(其中含10 mg/L 盐酸万古霉素, 2.5 mg/L 多黏菌素B, 2 mg/L 两性霉素B及75 mL/L 脱纤维羊血)制平板, 将已研磨的匀浆用布氏肉汤分别稀释10、100、1 000倍, 取稀释液100 μL涂平板, 37 °C微需氧(50 mL/L O₂, 100 mL/L CO₂, 850 mL/L N₂)培养72 h。挑取呈细小圆形、半透明菌落, 经Gram染色后, 于油镜下观察菌体形态。从细菌培养平板上挑取呈细小圆形、半透明菌落, 用胃H pylori 快速检测试剂盒HPUT-H104(购自福建三强公司)检测, 能使试剂变为橙色的为阳性菌落^[18]。

胃黏膜经常规甲醛液体固定、梯度乙醇脱水、二甲苯脱乙醇、石蜡包埋切片、脱蜡、梯度复水、HE染色、梯度脱水、二甲苯透明及树脂封片后, 于光学显微镜下观察胃组织病理改变^[19]。动物眼球取血后, 将血液收集于1.5 mL离心管中, 常温下静置, 离心收集血清。用酶联免疫黏附试验(ELISA)测定血清中抗H pylori IgG抗体。

2 结果

2.1 细菌分离培养及尿素酶试验 在H pylori 感染后4、12及24 wk, 分批处死实验组的小鼠, 在胃组织中分离到典型H pylori, 平板培养的菌落呈圆形、凸起、光滑、半透明状。经Gram染色后于油镜下观察, 其中被染成红色、呈弯曲螺旋状的菌体, 与在实验室中培养的H pylori 标准菌株形态一致(图1)。对照组的胃黏膜匀浆液中则未分离到典型H pylori 菌落。通过进一步的细菌分

离培养, 对每块平板进行计数, 结果见表1。实验组及对照组小鼠的胃黏膜匀浆液均进行快速尿素酶检测, 结果见表2。

2.2 病理组织学检查 H pylori 实验组于12wk 在胃门前区小弯处可见有慢性炎症变化, 即有多量的淋巴细胞和嗜中性粒细胞浸润。24wk 时炎症更加严重, 浸润细胞主要是淋巴细胞。而对照组小鼠胃则未发现有炎性细胞浸润现象(图2)。

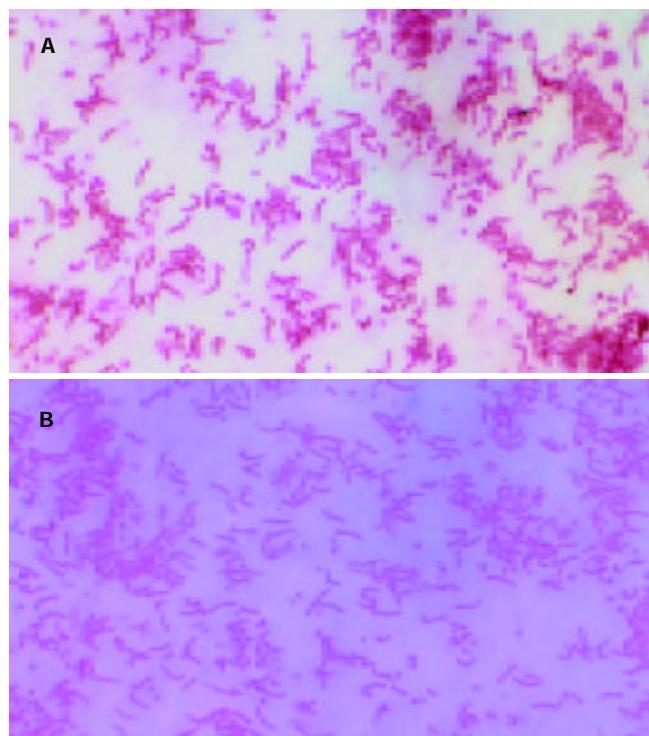


图1 H pylori 感染后4 wk 小鼠胃部分离的H pylori SS1 菌体形态. A: 标准株; B: 分离株。

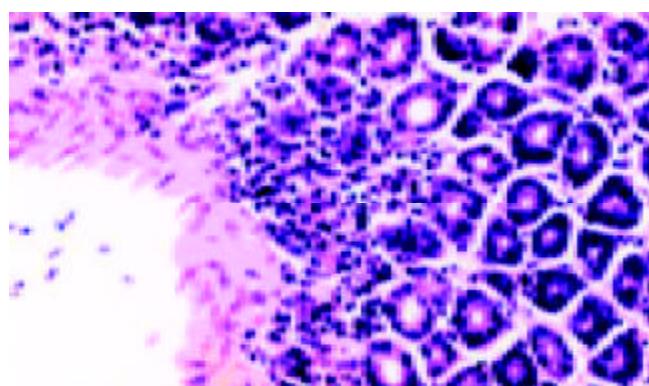


图2 H pylori 感染24 wk 小鼠胃黏膜见大量淋巴细胞浸润(HE染色)×400。

表1 菌落计数结果

小鼠类型	Balb/c			C57BL/6			
	处理时间	处死小鼠只数	检出阳性只数	H pylori 菌数 / 克胃组织(mean±SD)	处死小鼠只数	检出阳性只数	H pylori 菌数 / 克胃组织(mean±SD)
4 wk	7	6		1.44 × 10 ⁶ ± 19.48	7	5	1.12 × 10 ⁶ ± 18.33
12 wk	7	7		1.25 × 10 ⁶ ± 25.61	7	6	1.53 × 10 ⁶ ± 32.41
24 wk	7	7		1.03 × 10 ⁶ ± 57.22	7	6	1.84 × 10 ⁶ ± 26.75

表2 H pylori 感染小鼠尿素酶检测结果

处理时间(wk)	尿素酶检测阳性(%)	
	Balb/c	C57BL/6
4	85.7	71.4
12	100	85.7
24	100	85.7

2.3 血清抗体测定 采用ELISA法检测小鼠血清中的抗H pylori IgG抗体水平。其中抗原包被液为H pylori超声粉碎抗原,浓度为10 mg/L,每孔各加100 μL。以对照组小鼠的血清作为对照,一抗进行了1:1 000倍稀释,二抗为HRP-兔抗鼠IgG抗体。在各时间点H pylori实验组血清中抗H pylori的IgG抗体水平明显高于对照组($P < 0.01$),说明小鼠已感染了H pylori(表3)。

表3 H pylori 实验小鼠血清中抗H pylori IgG水平(mean±SD)

t /wk	对照组	H pylori 实验组
4	0.019 ± 0.004	0.080 ± 0.021
12	0.027 ± 0.017	0.102 ± 0.024
24	0.022 ± 0.013	0.113 ± 0.022

3 讨论

幽门螺杆菌(Helicobacter pylori, Hp)作为各种慢性胃病、慢性胃炎、消化性溃疡的重要病因,已得到公认。近年来世界各国对H pylori感染的免疫预防和免疫治疗的研究越来越重视,而良好的动物感染模型的建立是该研究的前提基础。尽管恒河猴、平顶猴、蒙古沙鼠及雪貂等是H pylori的天然宿主,但由于受价格贵、数量少、饲养条件高等因素限制,不适宜于做理想的动物模型研究^[20-23]。迄今只有少数动物感染成功。

虽然国外采用某些实验动物如悉生小猪、胸腺机能正常或异常的无菌小鼠等做H pylori动物模型,但这些动物不易大量获得,也不能用于免疫学研究^[24-25]。目前国际上公认的比较成熟的动物模型为Lee et al^[26]所建立的Helicobacter felis(Hf)小鼠模型,但不能直接反应H pylori疫苗的效果,不完全适用于H pylori疫苗免疫效果的评价。Karita et al^[27]也曾建立了H pylori的裸鼠感染模型,而裸鼠本身免疫功能异常,无法反映正常的免疫反应过程。国内有诸多研究人员用不同的动物进行了感染模型的研究,常用的有狗、大鼠、沙鼠等,但感染率不是很理想,并且感染的稳定性和重复性尚存在不同程度的问题^[28-30]。

小鼠体型小、易繁殖、来源便利、属系多样,是作为动物模型的较好材料^[31-35]。我们经实验研究表明,采用H pylori SS1菌株能成功感染Balb/c和C57BL/6小鼠,H pylori实验组胃黏膜匀浆液尿素酶检测试验结果:H pylori感染4 wk时Balb/c组阳性率为85.7%,C57BL/6组阳性率为71.4%,H pylori感染12 wk及24 wk

时,Balb/c组阳性率均为100%,C57BL/6组阳性率平均为85.7%,并且能从感染小鼠分离到与H pylori标准菌株形态一致的菌落;而对照组小鼠尿素酶检测为阴性,且不能分离到典型H pylori菌落。实验组动物胃窦部及胃体部均出现了轻度至中度慢性活动性胃炎变化,而对照动物均未见炎症变化。血清学IgG检查表明,实验组动物产生了抗H pylori血清抗体,而对照动物血清抗体阴性。这里应指出的是,在我们的实验中也出现了个别小鼠细菌分离阴性而血清抗体IgG检测呈阳性的情况,我们初步分析原因为,由于该鼠也同样经过了用H pylori菌液灌胃三次的实验步骤,即至少说也经历了外源细菌的感染过程,因而也产生了抗H pylori抗体。只不过可能由于个体差异或其他原因,H pylori在其胃内没有能够长期定植和繁殖,或者是由于定植密度太低而未能分离检测到。

以上结果表明,我们已成功建立了H pylori的小鼠动物模型,这对于H pylori的致病机制研究、疫苗开发研制等具有重要意义。

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