

肝移植缺血再灌注后Kupffer细胞CD14的表达机制

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CD14 expression in Kupffer cells of ischemia-reperfusion injury after rat liver transplantation

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Abstract

AIM: To study the expression of lipopolysaccharide receptor CD14 mRNA and protein in Kupffer cells and its role in ischemia-reperfusion injury (IRI) in rat liver graft.

METHODS: The Kupffer cells were isolated and divided into control, ischemia-reperfusion (IR), and anti CD14 antibody groups. The CD14 mRNA, CD14 protein, nuclear factor kappa B (NF- κ B) activity, and TNF- α and IL-1 level in the culture supernatant were measured.

RESULTS: The CD14 mRNA, and protein in IR group were significantly higher than those in control group (mRNA 1.28 ± 0.12 vs 0.42 ± 0.02 , protein 23.7 ± 2.36 vs 6.3 ± 1.27 , $P < 0.01$). The NF- κ B activity, TNF- α and IL-1 level in IR group were significantly higher than those in control group (NF- κ B 2.79 ± 0.48 vs 0.27 ± 0.01 , TNF- α 205.9 ± 12.04 ng/L vs 57.4 ± 4.35 ng/L, IL-1 176.8 ± 8.94 ng/L vs 37.6 ± 3.47 ng/L, $P < 0.01$), and they greatly decreased after anti-CD14 antibody treatment compared with IR group (NF- κ B 1.34 ± 0.24 vs 2.79 ± 0.48 , TNF- α 129.6 ± 6.48 ng/L vs 205.9 ± 12.04 ng/L, IL-1 103.4 ± 5.74 ng/L vs 176.8 ± 8.94 ng/L, $P < 0.05$), but still significantly higher than those in control group (NF- κ B 1.34 ± 0.24 vs 0.27 ± 0.01 , TNF- α 129.6 ± 6.48 ng/L vs 57.4 ± 4.35 ng/L, IL-1 103.4 ± 5.74 ng/L vs 37.6 ± 3.47 ng/L, $P < 0.01$).

CONCLUSION: LPS following IR can up-regulate the ex-

pression of CD14 mRNA and protein in Kupffer cells, and subsequently activate NF- κ B to produce cytokines. But other signal transduction pathways might also participate in the NF- κ B activation and IRI.

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摘要

目的: 研究大鼠肝移植缺血再灌注后Kupffer细胞脂多糖受体CD14表达即其参与缺血再灌注损伤的机制。

方法: 分离培养大鼠Kupffer细胞, 分为正常对照组, 肝移植缺血再灌注组, 抗CD14抗体组, 检测Kupffer细胞CD14mRNA、膜蛋白表达, 核转录因子 κ B活性、以及培养上清TNF- α 、IL-1的分泌量。

结果: 再灌注后Kupffer细胞CD14mRNA和膜蛋白表达明显高于对照组(mRNA 1.28 ± 0.12 vs 0.42 ± 0.02 ; 膜蛋白 23.7 ± 2.36 vs 6.3 ± 1.27 , $P < 0.01$); 再灌注后核转录因子 κ B活性、培养上清TNF- α 、IL-1表达量明显高于对照组(NF- κ B 2.79 ± 0.48 vs 0.27 ± 0.01 ; TNF- α 205.9 ± 12.04 ng/L vs 57.4 ± 4.35 ng/L; IL-1 176.8 ± 8.94 ng/L vs 37.6 ± 3.47 ng/L, $P < 0.01$); 用抗CD14抗体后NF- κ B活性、TNF- α 、IL-1表达量与再灌注相比明显下降(NF- κ B 1.34 ± 0.24 vs 2.79 ± 0.48 ; TNF- α 129.6 ± 6.48 ng/L vs 205.9 ± 12.04 ng/L; IL-1 103.4 ± 5.74 ng/L vs 176.8 ± 8.94 ng/L; $P < 0.05$), 但仍然高于对照组(NF- κ B 1.34 ± 0.24 vs 0.27 ± 0.01 ; TNF- α 129.6 ± 6.48 ng/L vs 57.4 ± 4.35 ng/L; IL-1 103.4 ± 5.74 ng/L vs 37.6 ± 3.47 ng/L, $P < 0.01$)。

结论: 缺血再灌注时LPS能够上调Kupffer细胞CD14表达, 激活NF- κ B, 启动细胞因子的转录和分泌, 但尚存在除CD14以外的其他信号途径参与了NF- κ B的激活和缺血再灌注损伤。

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0 引言

脂多糖(LPS)受体CD14是存在于单核-巨噬细胞表面的一种膜蛋白, 主要功能是识别LPS-LBP(脂多糖结合蛋白)复合物^[1-3]. LPS-LBP与CD14结合后, 促使单核-

巨噬细胞活化并释放多种前炎症因子，在内毒素血症、酒精性肝病、肝硬化等疾病中扮演重要角色^[4-8]。Kupffer细胞作为肝脏内一种独特的单核-巨噬细胞是否也表达CD14，CD14是否参与肝移植缺血再灌注损伤(ischemia-reperfusion injury, IRI)，引起IRI的细胞因子是否由CD14信号传导通路产生，这些问题到目前为止均未得到解决。我们通过分离培养肝移植缺血再灌注(IR)后的Kupffer细胞，观察CD14mRNA和蛋白的表达，CD14胞内信号传导途径中NF-κB活性以及细胞因子的产生情况，探讨其引起IRI的确切机制。

1 材料和方法

1.1 材料 实验分三组(每组30例)，对照组，取正常大鼠Kupffer细胞为实验对象；再灌注(IR)组，取再灌注后1 h Kupffer细胞为研究对象；抗CD14抗体组，取再灌注后1 h Kupffer细胞，并在培养液中加入CD14抗体0.2 mL共同培养12 h。健康♂Wistar大鼠，质量210-250 g(重庆医科大学实验动物中心提供)。参照Peng et al^[9]的改进Kamada袖套法，建立大鼠原位肝移植动物模型。供体手术：游离肝周韧带和血管，切取供肝，在水浴中完成门静脉、肝下下腔静脉袖套准备，胆管内支架插管。受体手术：切除原肝，原位植入供肝，先完成肝上下腔静脉连续缝合，门静脉袖套吻合，结束无肝期，以此时作为再灌注开始。完成肝下下腔静脉袖套吻合和胆道重建。再灌注后1 h，参照Gong et al介绍的胶原酶肝脏原位灌注法分离Kupffer细胞^[3]。经门静脉插管，50 g/L IV型胶原酶(Sigma)体外循环灌注消化肝脏，不连续Percol(Pharmacia)密度梯度离心分离Kupffer细胞，用含100 mL/L小牛血清的RPMI1640培养液，37 °C，50 mL/L CO₂培养6 h后，洗去未贴壁细胞，重悬贴壁细胞，调整细胞浓度为1×10⁶待用。测其纯度大于90%，活力大于95%。

1.2 方法

1.2.1 Kupffer细胞CD14 mRNA检测 采用Trizol试剂盒(life Technologies)提取Kupffer细胞总RNA，取0.5 mL RNA产物，用RT-PCR试剂盒(Roche)将其逆转录为互补DNA(cDNA)，-70 °C保存待用。以β-actin作为内参对照进行PCR反应(表1)，CD14以及β-actin引物由上海生工合成。PCR循环条件为：94 °C 1 min, 58 °C 1 min, 72 °C 1 min, 30个循环，72 °C延长7 min。用15 g/L琼脂糖凝胶电泳PCR产物，EB染色，凝胶成像系统和图像分析系统观察并半定量计算PCR产物的相对表达量，结果以CD14/β-actin的灰度比值表示。

1.2.2 Kupffer细胞CD14蛋白的检测 采用Western blot检测Kupffer细胞CD14蛋白的表达。Kupffer细胞蛋白提取物50 μg用100 g/L SDS-PAGE进行电泳分离，电泳后的蛋白质转移至硝酸纤维膜4 °C过夜，20 g/L脱脂奶粉封闭1 h，与抗CD14多克隆抗体(一抗，Santa Cruz)反应2 h，PBST洗涤3次去除封闭液和一抗，与辣根过

氧化酶标记的二抗(生物晶美公司)反应2 h，PBST洗涤3次去除二抗。最后加入增强化学发光剂自显影，凝胶成像系统进行密度扫描，图像分析软件分析蛋白区带，CD14蛋白表达量用蛋白区带积分吸光度表示。

表1 RT-PCR检测CD14引物设计

	引物序列	长度(bp)
CD14	5' CTC AAC CCT AGA GCG TTT CT 3'	267
	5' CAG GAT TG TC AGA CAG GT CT 3'	
β-actin	5' ATC ATG TTG AGA CAC CCT CAACA 3'	300
	5' CAT CT CTT GCT CGA AGTCCA 3'	

1.2.3 Kupffer细胞NF-κB活性检测 应用凝胶迁移变动分析(EMSA)检测Kupffer细胞NF-κB活性。提取KC细胞核蛋白，NF-κB寡核苷酸探针序列为5' GCC TCC AAT GTT CGC GAA CTT TCG 3'(Santa Cruz产品)，³²P标记，进行凝胶阻滞分析，放射自显影，测定每条电泳带后带的吸光度值，所得结果表示NF-κB的相对活性。

1.2.4 细胞培养上清液TNF-α和IL-1检测 采用ELISA检测试剂盒(Sigma)测定上清液TNF-α和IL-1(操作步骤参见说明书)。

统计学处理 实验数据以均数±标准差(mean±SD)表示，用SPSS9.0进行统计分析，以P<0.05为差别有显著性。

2 结果

2.1 Kupffer细胞CD14 mRNA表达 对照组有微量CD14 mRNA表达，再灌注后CD14 mRNA表达明显升高(CD14/β-actin 1.28±0.12 vs 0.42±0.02, P<0.01)。

2.2 Kupffer细胞CD14蛋白表达 再灌注后Kupffer细胞CD14蛋白明显表达，而对照组仅有微量的CD14蛋白表达(23.7±2.36 vs 6.3±1.27, P<0.01, 图1)。

2.3 Kupffer细胞NF-κB活性改变 再灌注后NF-κB活性明显高于对照组(2.79±0.48 vs 0.27±0.01, P<0.01)，应用抗CD14抗体后，NF-κB活性与IR相比显著降低(1.34±0.24 vs 2.79±0.48, P<0.05)，但仍然高于对照组(1.34±0.24 vs 0.27±0.01, P<0.01, 图2)。

2.4 TNF-α和IL-1表达量 IR组和抗CD14治疗组TNF-α和IL-1均明显高于对照组(P<0.01)，但抗CD14治疗组又明显低于IR组(P<0.05, 图3)。

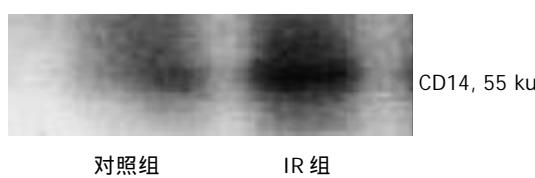


图1 CD14蛋白Western blot检测结果。

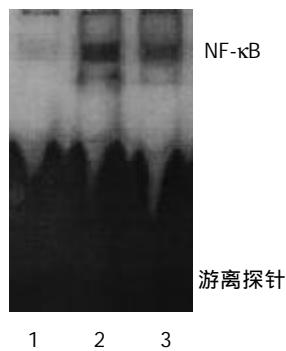


图2 NF-κB EMSA放射自显影图像. 1: 对照组; 2: IR组; 3: 抗CD14组.

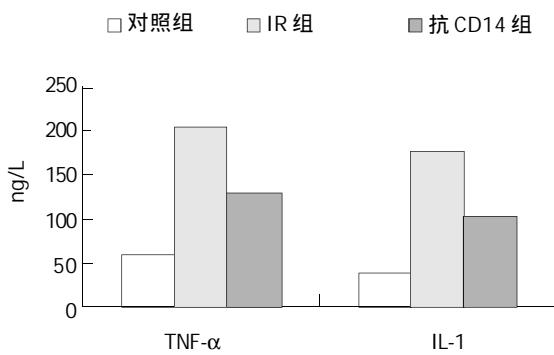


图3 Kupffer 细胞培养上清中 TNF- α 和 IL-1 分泌量.

3 讨论

CD14在介导内毒素引起的单核巨噬细胞活化过程中起启动作用^[2, 10-12], Kupffer 细胞是肝移植缺血再灌注损伤的主要参与者^[13-16], 因此研究 Kupffer 细胞上 CD14 的表达以及其后的胞内信号传导途径, 对于阐明肠道内毒素激活 Kupffer 细胞引起缺血再灌注损伤的机制意义重大^[17-20]. 我们发现, 正常处于静息状态下的 Kupffer 细胞有少量的 CD14 表达, 这可能对于维持正常 Kupffer 细胞功能, 使之处于应激状态是必需的^[15, 21-23]. 但再灌注后, 大量内毒素由门静脉入肝, 立即引起 CD14 mRNA 以及蛋白的表达大量增多, 启动 Kupffer 细胞的激活^[24-27].

缺血再灌注损伤的最终形式表现为大量有害细胞因子(如 TNF- α 、IL-1 等)对移植物的攻击和损伤^[28-31]. Kupffer 细胞 CD14 表达增加是否与细胞因子分泌增多存在因果关系? 我们通过研究 CD14 以后的胞内信号传导途径, 特别是处于 CD14 与细胞因子之间的 NF-κB 的活性变化, 阐明了他们之间的联系. 再灌注后 NF-κB 活性明显高于对照, TNF- α 等细胞因子的产生也较对照明显升高. 上述一系列连续的变化过程表明, LPS-CD14-NF-κB-细胞因子是存在于再灌注损伤中的一条重要信号传导途径, CD14 对其下游的 NF-κB 活化及细胞因子的产生存在必然关系. 但我们同时也发现, CD14 并非激活 NF-κB 唯一上游信号, 通过用抗 CD14 抗体阻断 CD14 功能后, 发现 NF-κB 活性以及细胞因子的产生有所降低, 但仍然高于正常, 证明除 CD14 途径外, 尚有其他信号途径可以激活 NF-κB.

总之, 我们的研究表明 Kupffer 有 CD14 基因以及膜蛋白的表达, 缺血再灌注后, CD14 表达增强, 内毒素通过与 CD14 的结合, 启动 Kupffer 细胞激活的信号传导, 信号通过多级酶联反应由胞外传到胞内, 激活 NF-κB, 启动 TNF- α 等细胞因子的转录和分泌, 最终造成对移植物的攻击和损害. 但尚需进一步研究 CD14 至 NF-κB 之间的准确信号传导过程, 以及除 CD14 以外的其他激活 NF-κB 的信号途径, 才能完整阐明参与缺血再灌注损伤的整个信号传导通路.

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