

银杏叶类黄酮对人胃癌细胞 BGC823 体外的增殖抑制作用

张凤, 杨桂文, 张金凤, 安利国

张凤, 杨桂文, 张金凤, 安利国, 山东师范大学生命科学学院动物抗性重点实验室, 山东省济南市 250014
 通讯作者: 安利国, 250014, 山东省济南市文化东路88号, 山东师范大学生命科学学院. anlg@sdnu.edu.cn
 电话: 0531-86180143
 收稿日期: 2005-06-28 接受日期: 2005-07-16

Inhibitory effects of *Ginkgo biloba* leaf flavonoids on proliferation of human gastric cancer cell line BGC823 *in vitro*

Feng Zhang, Gui-Wen Yang, Jin-Feng Zhang, Li-Guo An

Feng Zhang, Gui-Wen Yang, Jin-Feng Zhang, Li-Guo An, Key Laboratory of Animal Resistance, College of Life Sciences, Shandong Normal University, Jinan 250014, Shandong Province, China
 Correspondence to: Li-Guo An, College of Life Sciences, Shandong Normal University, 88 Wenhua East Road, Jinan 250014, Shandong Province, China. anlg@sdnu.edu.cn

Received: 2005-06-28 Accepted: 2005-07-16

Abstract

AIM: To extract the flavonoids from *Ginkgo biloba* leaf, and to investigate its inhibitory effects on the proliferation of human gastric cancer cell line BGC823 cultured *in vitro*.

METHODS: Ethanol (700 mL/L) was used to extract the flavonoids from the leaf of *Ginkgo biloba*. Three wavelength spectrophotometry was used to determine the content of flavonoids in the extracts. Human gastric cancer cells BGC823 cultured *in vitro* were treated with different concentrations of the flavonoids, and then the proliferation of the cells was detected by MTT assay and flow cytometry.

RESULTS: The content of flavonoids in the extracts was 140 mg/g. The flavonoids from *Ginkgo biloba* leaf inhibited the proliferation of BGC823 cells in a dose-dependent manner. The rate of cells in S phase was notably increased as compared with that in the controls ($42.17 \pm 0.50\% vs 32.13 \pm 0.45\%, P = 0.001$), and the apoptotic rate of the cells was also increased ($4.10 \pm 0.03\% vs 2.21 \pm 0.01\%, P = 0.002$).

CONCLUSION: *Ginkgo biloba* leaf flavonoids can inhibit the proliferation of human gastric cancer cell line BGC823 by affecting the cycle the cells.

Key Words: *Ginkgo biloba* leaf flavonoids; Gastric cancer

Zhang F, Yang GW, Zhang JF, An LG. Inhibitory effects of *Ginkgo biloba* leaf flavonoids on proliferation of human gastric cancer cell line BGC823 *in vitro*. *Shijie Huaren Xiaohua Zazhi* 2005;13(21):2627-2629

摘要

目的: 提取并测定银杏叶类黄酮(芦丁)的含量; 探讨银杏叶类黄酮对体外培养的人胃癌细胞BGC823的增殖抑制作用.

方法: 乙醇法(700 mL/L)提取银杏叶类黄酮; 三波长分光光度法测定提取物中类黄酮(芦丁)的含量; MTT法及流式细胞技术观察提取物对人胃癌细胞BGC823的增殖抑制作用.

结果: 提取物中类黄酮(芦丁)含量为140 mg/g. MTT法证实银杏叶类黄酮对人胃癌细胞BGC823增殖有抑制作用, 且呈剂量依赖效应. 流式细胞术分析表明银杏叶类黄酮将胃癌细胞BGC823的生长周期阻滞于S期, 与对照组相比明显增加($42.17 \pm 0.50\% vs 32.13 \pm 0.45\%, P = 0.001$), 凋亡细胞数与对照组相比也显著增加($4.10 \pm 0.03\% vs 2.21 \pm 0.01\%, P = 0.002$).

结论: 银杏叶类黄酮对人胃癌细胞BGC823增殖有抑制作用, 并能阻抑细胞周期进程, 诱导细胞凋亡.

关键词: 银杏叶类黄酮; 胃癌

张凤, 杨桂文, 张金凤, 安利国. 银杏叶类黄酮对人胃癌细胞BGC823体外的增殖抑制作用. 世界华人消化杂志 2005;13(21):2627-2629
<http://www.wjgnet.com/1009-3079/13/2627.asp>

0 引言

银杏(*Ginkgo biloba* L.)又名白果树, 银杏叶提取物(GBE)主要药用成分之一为类黄酮^[1-3], 用于对心血管、动脉硬化、高血压等疾病的治疗, 具有独特药理活性和巨大的临床应用价值^[4-9]. Lee *et al*^[10]报道银杏提取物中的双黄酮—银杏黄素或异银杏黄素10 μmol/L能抑制ConA或LPS诱导的淋巴细胞增殖, Chao *et al*^[11]研究表明, 银杏提取物EGb761能够抑制肝癌细胞HepG2和Hep3B的增殖, Kim *et al*^[12]研究显示, 银杏提取物EGb761能够激活caspase-3诱导口腔牙槽癌细胞凋亡. 据此我们推测银杏叶提取物也许能够通过抑制细胞增殖从而抑制胃癌细胞的生长, 在胃癌治疗中发挥有益作用. 我们研究银杏叶提

取物中类黄酮含量并探讨其对胃癌细胞BGC823细胞增殖的抑制作用,为进一步开发银杏这一我国特有植物物种资源的应用价值提供基础资料。

1 材料和方法

1.1 材料 银杏叶; BGC823细胞株; RPMI-1640培养基(Gibco); 小牛血清(杭州四季青公司); 二甲基亚砜(DMSO); 噻唑蓝(MTT)(Sigma); 芦丁标准品(中国药品检验所)。类黄酮的提取(700 mL/L乙醇法)参照秦雪莲等^[13-15]的方法,称取银杏叶干粉10 g,加700 mL/L乙醇400 mL,混匀,40℃超声振荡50 min,8 000 g离心20 min,收集上清。滤渣再加700 mL/L乙醇300 mL,混匀,40℃超声振荡30 min,8 000 g离心20 min,收集上清。滤渣再加700 mL/L乙醇100 mL,混匀,40℃超声振荡20 min,8 000 g离心20 min,收集上清。将所得的3次上清液合并,减压浓缩并将浓缩液置于硫酸纸上,40℃干燥箱中干燥,所得干燥品即为银杏叶粗黄酮。称取芦丁标准品5 mg溶于水中并定容于100 mL容量瓶中,即为芦丁母液;取母液0.5,1.0,1.5,2.0,2.5,3.0,3.5 mL于容量瓶中,每一容量瓶用水加至3.5 mL,再用10 g/L AlCl₃定容至5 mL;测定 $\lambda_1=470$ nm, $\lambda_2=420$ nm, $\lambda_3=370$ nm三波长处的吸光度A值。按公式 $\Delta A=A_2-\{[(\lambda_2-\lambda_3)A_1+(\lambda_1-\lambda_2)A_3]/(\lambda_1-\lambda_3)\}$ 计算 ΔA ,根据 ΔA 与浓度关系绘制标准曲线。样品中类黄酮含量的测定:称取银杏叶粗黄酮5 mg溶于水中,并定容于5 mL容量瓶中,即为粗黄酮溶液,取此液2.5 mL于5 mL容量瓶中,用水加至3.5 mL,用10 g/L AlCl₃定容至刻度,测定 $\lambda_1=470$ nm, $\lambda_2=420$ nm, $\lambda_3=370$ nm三波长处的吸光度A值,计算样品中黄酮含量。10 g银杏叶中提取得到类黄酮1.95 g,得率为19.5%。根据 ΔA 与浓度关系绘制标准曲线,回归方程为:Y=31.096x+0.013 4, R²=0.999 7。样品中类黄酮含量计算可知1 mg提取物中含黄酮140 μg,进而根据提取物得率计算银杏叶中类黄酮含量为2.73%。

1.2 方法 取对数生长期的胃癌细胞BGC823细胞以 5×10^7 /L浓度接种于96孔培养板,每孔100 μL,37℃,50 mL/L CO₂培养箱中培养。24 h后分别接入不同浓度的无菌的银杏叶类黄酮溶液100 μL,使最终浓度分别为60,80,100,150,200,250 g/L,各组设8个重复孔,对照组加100 μL培养液,在37℃,50 mL/L CO₂条件下继续培养48 h。实验终止前加入新配制的5 g/L的MTT溶液20 μL,混匀,再继续培养4 h。弃去上清,每孔加DMSO 200 μL,充分振荡30 min,溶解MTT沉淀物,490 nm测定每孔的吸光度A值^[16-19]。计算抑制率:抑制率=(1-实验组A值/对照组A值)×100%。另将 5×10^7 /L细胞接种于2个6 cm培养皿中,24 h后加入无菌银杏叶类黄酮溶液,使其终浓度为150 g/L,对照组加等量的培养液。继续培养48 h,终止培养后制成单细胞悬液,调整细胞浓度为 1×10^9 /L,离心,用同体积700 mL/L的冷乙醇固定1 h,离心,PBS洗涤2次,

将细胞悬液和碘化丙啶(PI)等体积混合,4℃放置30 min,放入流式细胞仪样品室,488 nm检测细胞周期和细胞凋亡的情况^[20,21]。

统计学处理 用SPSS 10.0统计学软件和t检验处理数据,所有数据均用mean±SD表示。

2 结果

2.1 银杏叶类黄酮对胃癌细胞BGC823的抑制作用 银杏叶类黄酮对肿瘤细胞BGC823增殖有抑制作用,并呈现明显的剂量依赖效应(表1)。

表1 银杏叶类黄酮对胃癌细胞增殖的抑制作用

黄酮(mg/L)	吸光度(A)	抑制率(%)
0	0.675 ± 0.096	-
60	0.596 ± 0.084 ^b	11.5 ± 1.4
80	0.555 ± 0.164 ^a	17.4 ± 6.6
100	0.484 ± 0.057 ^b	28.1 ± 2.4
150	0.361 ± 0.094 ^b	46.4 ± 2.8
200	0.168 ± 0.022 ^b	75.2 ± 0.5
250	0.096 ± 0.013 ^b	85.9 ± 0.4

^aP<0.05, ^bP<0.01 vs 0 mg/L。

2.2 流式细胞仪分析结果 银杏叶类黄酮使胃癌细胞BGC823的细胞周期滞留于S期,并且能够诱导细胞凋亡(表2)。

表2 流式细胞仪分析的BGC823细胞周期分布 (mean ± SD %)

银杏叶黄酮类	G ₀ -G ₁ 期	S期	G ₂ -M期	凋亡率
对照	58.45 ± 0.35	32.13 ± 0.45	9.41 ± 0.14	2.21 ± 0.11
150 mg/L	57.72 ± 0.45	42.17 ± 0.50 ^b	0.11 ± 0.08 ^b	4.10 ± 0.03 ^b

^bP<0.01 vs 对照。

3 讨论

在类黄酮化合物的紫外光谱吸收中,主要吸收带是由304-350 nm的吸收带I和240-280 nm的吸收带II组成。因为银杏叶类黄酮提取物中其它成分在带I和带II范围内也有一定程度的吸收,会对类黄酮的测定产生干扰。加入铝盐使类黄酮与铝离子形成稳定的配合物,吸收带I会明显红移,同时吸光度也大大增加。因此,选择铝配合物显色体系来测定总黄酮的含量可以去除其他成分的干扰^[22,23],同时选择三波长一光谱法能够有效地消除吸收峰不对称给定量分析造成的影响,提高定量分析的准确度。通过数据分析可知银杏叶粗提取物中类黄酮含量为140 mg/g,银杏叶中类黄酮的含量约为2.73%。

通过MTT法观察银杏叶类黄酮对人胃癌细胞增殖的影响表明,银杏叶类黄酮对人胃癌细胞的增殖有抑制作用,而且存在明显的剂量依赖效应。我们在中效浓度附近选择了150 mg/L进一步用流式细胞仪分析其对胃癌细胞BGC823的作用,结果表明处理的胃癌细胞BGC823 S期细

胞比例明显增加(42.17 ± 0.50 vs $32.13 \pm 0.45, P = 0.001$), 并且能够诱导细胞凋亡(4.10 ± 0.03 vs $2.21 \pm 0.01, P = 0.002$), 这说明银杏叶类黄酮能够把胃癌细胞BGC823的细胞周期阻滞在S期, 阻碍其向G₂/M期的转换, 减缓其分裂的速度, 从而抑制其增殖并诱导细胞凋亡. 这与许多研究者对抗癌活性物质研究的实验结果类似^[24-28], 其诱导癌细胞凋亡的机制可能与p53和Fas有关^[29,30]. 然而, 银杏叶类黄酮如何通过阻抑细胞周期进程并诱导细胞凋亡起到抗肿瘤作用的至今还不清楚. 因此, 对于银杏叶类黄酮抗肿瘤的作用机制尚需进一步研究.

4 参考文献

- 1 黄沛力, 李建新, 张淑华, 王晖. 银杏叶不同有效成分抑制金属离子诱导人血浆低密度脂蛋白氧化修饰的比较研究. 中药材 2004; 27: 654-656
- 2 Gaudineau C, Beckerman R, Welbourn S, Auclair K. Inhibition of human P450 enzymes by multiple constituents of the *Ginkgo biloba* extract. *Biochem Biophys Res Commun* 2004; 318: 1072-1078
- 3 Nishida S, Satoh H. Comparative vasodilating actions among terpenoids and flavonoids contained in *Ginkgo biloba* extract. *Clin Chim Acta* 2004; 339: 129-133
- 4 Lin SJ, Yang TH, Chen YH, Chen JW, Kwok CF, Shiao MS, Chen YL. Effects of *Ginkgo biloba* extract on the proliferation of vascular smooth muscle cells *in vitro* and on intimal thickening and interleukin-1beta expression after balloon injury in cholesterol-fed rabbits *in vivo*. *J Cell Biochem* 2002; 85: 572-582
- 5 潘洪平. 银杏叶制剂药理作用和临床应用研究进展. 中国中药杂志 2005; 30: 93-96
- 6 Satoh H, Nishida S. Electropharmacological actions of *Ginkgo biloba* extract on vascular smooth and heart muscles. *Clin Chim Acta* 2004; 342: 13-22
- 7 Villasenor-Garcia MM, Lozoya X, Osuna-Torres L, Viveros-Paredes JM, Sandoval-Ramirez L, Puebla-Perez AM. Effect of *Ginkgo biloba* extract EGB 761 on the nonspecific and humoral immune responses in a hypothalamic-pituitary-adrenal axis activation model. *Int Immunopharmacol* 2004; 4: 1217-22
- 8 Yao ZX, Han Z, Drieu K, Papadopoulos V. *Ginkgo biloba* extract (Egb 761) inhibits beta-amyloid production by lowering free cholesterol levels. *J Nutr Biochem* 2004; 15: 749-56
- 9 朱贵月, 朱兴雷, 耿庆信, 张兴华, 邵建华. 冠心病患者外周血单核细胞清道夫受体活性变化及银杏叶提取物的干预作用. 中国中西医结合杂志 2004; 24: 1069-1072
- 10 Lee SJ, Choi JH, Son KH, Chang HW, Kang SS, Kim HP. Suppression of mouse lymphocyte proliferation *in vitro* by naturally-occurring biflavonoids. *Life Sci* 1995; 57: 551-558
- 11 Chao JC, Chu CC. Effects of *Ginkgo biloba* extract on cell proliferation and cytotoxicity in human hepatocellular carcinoma cells. *World J Gastroenterol* 2004; 10: 37-41
- 12 Kim KS, Rhee KH, Yoon JH, Lee JG, Lee JH, Yoo JB. *Ginkgo biloba* extract (EGB 761) induces apoptosis by the activation of caspase-3 in oral cavity cancer cells. *Oral Oncol* 2005; 41: 383-389
- 13 秦雪莲. 甜茶叶中黄酮类化合物提取条件研究. 四川化工与腐蚀控制 2003; 6: 13-15
- 14 刘心平, 徐文弟. 银杏叶黄酮提取工艺及对HeLa细胞Bcl-2 mRNA表达的影响. 哈尔滨医科大学学报 2005; 39: 250-252
- 15 侯冬岩, 回瑞华, 杨梅, 李铁纯, 刘晓媛, 朱永强. 绿茶及其饮料中总黄酮的分析. 分析试验室 2003; 22: 86-88
- 16 Huang C, Liu LY, Song TS, Ni L, Yang L, Hu XY, Hu JS, Song LP, Luo Y, Si LS. Apoptosis of pancreatic cancer BXPC-3 cells induced by indole-3-acetic acid in combination with horseradish peroxidase. *World J Gastroenterol* 2005; 11: 4519-4523
- 17 Wu K, Yuan LH, Xia W. Inhibitory effects of apigenin on the growth of gastric carcinoma SGC-7901 cells. *World J Gastroenterol* 2005; 11: 4461-4464
- 18 Wang D, Xiang DB, He YJ, Li ZP, Wu XH, Mou JH, Xiao HL, Zhang QH. Effect of caffeic acid phenethyl ester on proliferation and apoptosis of colorectal cancer cells *in vitro*. *World J Gastroenterol* 2005; 11: 4008-4012
- 19 Zhang R, Gong J, Wang H, Wang L. Bile salts inhibit growth and induce apoptosis of human esophageal cancer cell line. *World J Gastroenterol* 2005; 11: 5109-5116
- 20 Matsumoto K, Akao Y, Ohguchi K, Ito T, Tanaka T, Jinuma M, Nozawa Y. Xanthones induce cell-cycle arrest and apoptosis in human colon cancer DLD-1 cells. *Bioorg Med Chem* 2005
- 21 Manthey KC, Rodriguez-Melendez R, Hoi JT, Zempleni J. Riboflavin deficiency causes protein and DNA damage in HepG2 cells, triggering arrest in G1 phase of the cell cycle. *J Nutr Biochem* 2005
- 22 回瑞华, 侯冬岩, 关崇新, 刘晓媛. 三波长-光谱法测定沙棘果汁中黄酮的含量. 光谱学与光谱分析 2005; 25: 266-269
- 23 侯冬岩, 回瑞华, 杨梅, 刘晓媛, 关崇新. 苦丁茶中总黄酮的三波长-光法定量分析. 分析化学 2004; 32: 783-786
- 24 Lee HZ, Leung HW, Lai MY, Wu CH. Baicalein induced cell cycle arrest and apoptosis in human lung squamous carcinoma CH27 cells. *Anticancer Res* 2005; 25: 959-964
- 25 Zhang M, Chen H, Huang J, Li Z, Zhu C, Zhang S. Effect of lycium barbarum polysaccharide on human hepatoma QGY7703 cells: inhibition of proliferation and induction of apoptosis. *Life Sci* 2005; 76: 2115-2124
- 26 Espinosa A, Marchal JA, Aranega A, Gallo MA, Aiello S, Campos J. Antitumour properties of benzannelated seven-membered 5-fluorouracil derivatives and related open analogues. Molecular markers for apoptosis and cell cycle dysregulation. *Farmaco* 2005; 60: 91-97
- 27 Leung HW, Wu CH, Lin CH, Lee HZ. Luteolin induced DNA damage leading to human lung squamous carcinoma CH27 cell apoptosis. *Eur J Pharmacol* 2005; 508: 77-83
- 28 Iguchi T, Miyakawa Y, Yamamoto K, Kizaki M, Ikeda Y. Nitrogen-containing bisphosphonates induce S-phase cell cycle arrest and apoptosis of myeloma cells by activating MAPK pathway and inhibiting mevalonate pathway. *Cell Signal* 2003; 15: 719-727
- 29 Katayama K, Ueno M, Yamauchi H, Nagata T, Nakayama H, Doi K. Ethylnitrosourea induces neural progenitor cell apoptosis after S-phase accumulation in a p53-dependent manner. *Neurobiol Dis* 2005; 18: 218-225
- 30 N'cho M, Brahmi Z. Evidence that Fas-induced apoptosis leads to S phase arrest. *Hum Immunol* 2001; 62: 310-319