



丙型肝炎肝硬化CD56⁺T细胞和NK细胞数量及抗肿瘤活性

范荣山, 于德军, 孙德荣

■背景资料

通过流行病学调查研究, 慢性丙型肝炎发展为肝硬化和肝癌的比例较高, 其确切机制目前还不清楚。肝脏局部的免疫环境在肝癌的发生发展中的作用越来越受到人们的重视。肝脏中分布着大量天然和获得性的T细胞、NK细胞和CD56⁺T细胞(NKT), NKT同时表达T细胞受体和NK细胞受体, 是抗肿瘤早期起作用的效应细胞, 其重要性受到广泛的关注。

范荣山, 孙德荣, 大庆市第二医院内科 黑龙江省大庆市

163461

于德军, 大庆市第二医院检验科 黑龙江省大庆市 163461

通讯作者: 范荣山, 163461, 黑龙江省大庆市红岗区解放一街, 大庆市第二医院内科. rsfan519@163.com

电话: 0459-5202419

收稿日期: 2005-07-11 接受日期: 2005-08-11

productions of IFN- γ in the three groups were 7.4 ± 2.4, 3.2 ± 1.8 and 1.9 ± 0.5 μ g/L, respectively; the anti-tumor activities hepatic MNC in the three groups were 61.1% ± 17.1%, 59.2% ± 14.6%, and 26.7% ± 8.5%, respectively. For the above four groups of parameters, the changes in HC-induced cirrhosis patients was the most significant ($P < 0.05$).

CONCLUSION: The numbers and anti-tumor activities of CD56⁺T cells and NK cells are decreased in cirrhotic livers with HC.

Key Words: Hepatitis C; Liver cirrhosis; CD56⁺ T cell; Natural killer cell; Cell number; Activity

Fan RS, Yu DJ, Sun DR. Numbers and activities of CD56⁺ T cells and natural killer cells in cirrhotic livers with hepatitis C. Shijie Huaren Xiaohua Zazhi 2006;14(18):1836-1838

摘要

目的: 研究CD56⁺T细胞和NK细胞在丙型肝炎肝硬化时的数量及抗肿瘤细胞活性变化。

方法: 丙型肝炎肝硬化患者18例, 快速肝穿获得肝组织, 分离肝脏单个核细胞(MNC), 流式细胞仪分析CD56⁺T细胞及NK细胞数量, 肝脏MNC和外周血单个核细胞(PBMC)分别与IL-2混合培养, 检测IFN- γ 含量及抗肿瘤细胞活性。

结果: 健康志愿者 慢性丙型肝炎患者 丙型肝炎肝硬化患者肝脏MNC中CD56⁺T细胞数量百分比分别是20.4%± 6.2%, 11.2%± 3.1%和5.0%± 1.6%; NK细胞比例分别是31.1%± 9.7%, 31.6%± 8.3%和18.3%± 5.4%; 肝脏MNC的IFN- γ 产量分别是7.4± 2.4, 3.2± 1.8和1.9± 0.5 μ g/L; 抗肿瘤细胞的细胞毒性分别是61.1%± 17.1%, 59.2%± 14.6%, 26.7%± 8.5%。以上四组数据均以肝炎肝硬化患者变化最为显著($P < 0.05$)。

结论: 丙型肝炎肝硬化时肝脏CD56⁺T细胞和NK细胞数量及抗肿瘤细胞活性分别下降。

关键词: 丙型肝炎; 肝硬化; CD56⁺T细胞; NK细胞;

■相关报道

Kawarabayashi *et al*研究了手术得到的标本, 分析了肝癌和非肝癌患者肝脏中NK和NKT细胞数量及抗肿瘤活性; Ogasawara *et al*认为IFN- γ 是NK及NKT发挥抗肿瘤作用的重要的细胞因子。

Abstract

AIM: To study the numbers and anti-tumor activities of CD56⁺ T cells and natural killer (NK) cells in cirrhotic liver with hepatitis C (HC).

METHODS: Hepatic mononuclear cells (MNC) were isolated from liver specimens obtained from the patients ($n = 16$) with HC-induced cirrhosis by liver biopsy. In addition, the numbers of CD56⁺ T cells and natural killer cells were determined by flow cytometry. Liver MNC and peripheral blood mononuclear cells (PBMC) were co-cultured with the interleukin-2 (IL-2), respectively, and the production of interferon- γ (IFN- γ) and the antitumor activity were measured.

RESULTS: The percentages of CD56⁺ T cells among hepatic MNC in health individuals, HC and HC-induced cirrhosis patients were 20.4% ± 6.2%, 11.2% ± 3.1% and 5.0% ± 1.6%, respectively; the proportions of NK cells among liver MNC in the three groups were 31.1% ± 9.7%, 31.6% ± 8.3% and 18.3% ± 5.4%; the productions of IFN- γ in the three groups were 7.4 ± 2.4, 3.2 ± 1.8 and 1.9 ± 0.5 μ g/L; the anti-tumor activities of hepatic MNC in the three groups were 61.1% ± 17.1%, 59.2% ± 14.6%, 26.7% ± 8.5%. The changes in HC-induced cirrhosis patients were the most significant ($P < 0.05$).

■ 应用要点

改善肝脏局部的抗肿瘤免疫功能，可能成为预防肝炎肝硬化发生肝癌的有效措施。

^[13-14]	CD56 ⁺ T	CD56 ⁻ T	NK
HCV		IFN- γ , IL-2	IL-12, CD56 ⁻ T
	^[15] , IFN- γ		
MNC	CD56 ⁺ T	NK	
		CD56 ⁺ T	NK
IL-2	IL-12	,	IFN- γ , ^[16-17]
	,		
PBMC	CD56 ⁺ T	NK	IL-2 IL-12 , ^[18]
	CD56 ⁺ T	NK	IFN- γ , ,
		CD56 ⁺ T	NK

4 参考文献

- 1 王豪.丙型肝炎的流行病学与预防.中华肝脏病杂志
2003; 11: 366-367

2 邱大鹏,邱双健,吴志全,樊嘉,叶胜龙,余耀,周俭,蔡
晓燕.NKT细胞在肝癌组织中的分布状况与肝癌局部
免疫的研究.中国临床医学 2004; 11: 567-569

3 Kawachi Y, Watanabe H, Moroda T, Haga M, Imai
T, Hatakeyama K, Abo T. Self-reactive T cell clones
in a restricted population of interleukin-2 receptor
beta+ cells expressing intermediate levels of the T
cell receptor in the liver and other immune organs.
Eur J Immunol 1995; 25: 2272-2278

4 Kawachi Y, Arai K, Moroda T, Kawamura T,
Umezawa H, Naito M, Ohtsuka K, Hasegawa K,
Takahashi-Iwanaga H, Iwanaga T. Supportive
cellular elements for hepatic T cell differentiation:
T cells expressing intermediate levels of the T cell
receptor are cytotoxic against syngeneic hepatoma,
and are lost after hepatocyte damage. *Eur J Immunol*
1995; 25: 3452-3459

5 Lombard C, McKallip RJ, Hylemon PB, Nagarkatti
PS, Nagarkatti M. Fas Ligand-dependent and
-independent mechanisms of toxicity induced by
T cell lymphomas in lymphoid organs and in the
liver. *Clin Immunol* 2003; 109: 144-153

6 Miyaji C, Watanabe H, Osman Y, Kuwano Y, Abo T.
A comparison of proliferative response to IL-7 and
expression of IL-7 receptors in intermediate TCR
cells of the liver, spleen, and thymus. *Cell Immunol*
1996; 169: 159-165

7 Tavian M, Peault B. Embryonic development of the

- human hematopoietic system. *Int J Dev Biol* 2005; 49: 243-250

8 Watanabe H, Miyaji C, Seki S, Abo T. c-kit+ stem cells and thymocyte precursors in the livers of adult mice. *J Exp Med* 1996; 184: 687-693

9 Massa S, Balciunaite G, Ceredig R, Rolink AG. Critical role for c-kit (CD117) in T cell lineage commitment and early thymocyte development *in vitro*. *Eur J Immunol* 2006; 36: 526-532

10 Shimizu T, Bannai M, Kawamura H, Yamamoto S, Oya H, Maruyama S, Minagawa M, Kawamura T, Watanabe H, Hatakeyama K, Abo T. Organ specificity of c-kit+ lymphoid precursors in the liver, thymus, and bone marrow. *Eur J Haematol* 2000; 64: 416-425

11 Banerjee D, Liou HC, Sen R. c-Rel-dependent priming of naive T cells by inflammatory cytokines. *Immunity* 2005; 23: 445-458

12 Heink S, Ludwig D, Kloetzel PM, Kruger E. IFN-gamma-induced immune adaptation of the proteasome system is an accelerated and transient response. *Proc Natl Acad Sci USA* 2005; 102: 9241-9246

13 Schijns VE, Wierda CM, van Hoeij M, Horzinek MC. Exacerbated viral hepatitis in IFN-gamma receptor-deficient mice is not suppressed by IL-12. *J Immunol* 1996; 157: 815-821

14 Khan S, Zimmermann A, Basler M, Groettrup M, Hengel H. A cytomegalovirus inhibitor of gamma interferon signaling controls immunoproteasome induction. *J Virol* 2004; 78: 1831-1842

15 Kawarabayashi N, Seki S, Hatsuse K, Ohkawa T, Koike Y, Aihara T, Habu Y, Nakagawa R, Ami K, Hiraide H, Mochizuki H. Decrease of CD56(+)T cells and natural killer cells in cirrhotic livers with hepatitis C may be involved in their susceptibility to hepatocellular carcinoma. *Hepatology* 2000; 32: 962-969

16 Kenna T, Golden-Mason L, Porcelli SA, Koezuka Y, Hegarty JE, O'Farrelly C, Doherty DG. NKT cells from normal and tumor-bearing human livers are phenotypically and functionally distinct from murine NKT cells. *J Immunol* 2003; 171: 1775-1779

17 Ogasawara K, Takeda K, Hashimoto W, Satoh M, Okuyama R, Yanai N, Obinata M, Kumagai K, Takada H, Hiraide H, Seki S. Involvement of NK1+ T cells and their IFN-gamma production in the generalized Shwartzman reaction. *J Immunol* 1998; 160: 3522-3527

18 Satoh M, Seki S, Hashimoto W, Ogasawara K, Kobayashi T, Kumagai K, Matsuno S, Takeda K. Cytotoxic gammadelta or alphabeta T cells with a natural killer cell marker, CD56, induced from human peripheral blood lymphocytes by a combination of IL-12 and IL-2. *J Immunol* 1996; 157: 3886-3892

电编 张敏 编辑 潘伯荣